

# Molecular modeling study of intercalation complexes of tricyclic carboxamides with d(CCGGCGCCGG)<sub>2</sub> and d(CGCGAATTCGCG)<sub>2</sub>

Athanasia Varvaresou · Kriton Iakovou

Received: 30 May 2010 / Accepted: 29 October 2010 / Published online: 14 December 2010  
© Springer-Verlag 2010

**Abstract** Tricyclic dyes with different mesoatoms such as xanthenes (fluorescein, eosin) anthracenes and acridines (proflavine) approved by the Food and Drug Administration (FDA) for use in foods, pharmaceuticals and cosmetic preparations interact with DNA, and some of them do so through intercalation. Hyperchem 7.5, Spartan 04, Yasara 10.5.14 program packages and molecular modeling, molecular mechanics and dynamics techniques with the oligonucleotides d(CCGGCGCCGG)<sub>2</sub> and d(CGCGAATTCGCG)<sub>2</sub> were utilized in order to examine the mode of binding to DNA of a range of tricyclic carboxamides bearing N,N-dimethylaminoethyl side chain, i.e., 9-amino-DACA, anthracene, acridine-1-carboxamide, acridine-4-carboxamide (DACA), azacridine, phenazine, pyridoquinoxaline, oxopyridoquinoxaline, phenoxazine and xanthenone or N,N-dimethylaminobutyl moiety, i.e., phenazine and acridine. The bicyclic quinoline-8-carboxamide was also examined for comparison reasons. On the basis of our data, prerequisite for the interaction between protonated N,N-dimethylaminoethyl moiety and guanine is the formation of only one internal hydrogen bond between carboxamide and *peri* NH + in the case of 9-amino-DACA or *peri* N in the cases of DACA, azacridine, phenazine and pyridoquinoxaline. The presence of an additional internal hydrogen bond

between oxygen carboxamide and protonated N,N-dimethylamino group in the cases of tricyclic systems bearing *peri* NH (phenoxazine) or O (xanthenone) group, prevents the interaction between side chain and guanine. Also, the formation of one internal hydrogen bond between oxygen carboxamide and protonated N,N-dimethylamino group inhibits the interaction between side chain and guanine in the case of acridine-1-carboxamide. Our findings are in accordance with previously reported results obtained from the kinetic studies of the binding of acridine and related tricyclic carboxamides to DNA.

**Keywords** DNA intercalation · Molecular dynamic simulations · Molecular mechanics · Tricyclic dyes

## Introduction

Tricyclic dyes with different mesoatoms such as xanthenes (fluorescein, eosin) anthracenes and acridines (proflavine) approved by the Food and Drug Administration (FDA) for use in foods, pharmaceuticals and cosmetic preparations interact with DNA, and some of them do so through intercalation [1–4].

The intercalation process reflects the ability of a planar aromatic or heteroaromatic system to become inserted between adjacent base pairs of a DNA molecule without disturbing the overall stacking pattern [5].

The acridine-4-carboxamides are a series of DNA intercalating topoisomerase poisons, developed by Denny and colleagues, that show remarkable variation in antitumor activity for minimal changes in molecular structure [6]. Some are potent topoisomerase poisons with widespread antitumor efficacy, for example, N-[2-(dimethylamino)ethyl]acridine-4-carboxamide (DACA), which is a DNA-

Kriton Iakovou contributed equally to this work.

A. Varvaresou (✉)  
Laboratory of Cosmetology, Department of Aesthetics and  
Cosmetology, Technological Educational Institution of Athens,  
Ag. Spyridona, Egaleo,  
12 210, Athens, Greece  
e-mail: varvaresou@pharm.uoa.gr

K. Iakovou  
Ministry of Health and Social Solidarity,  
Aristotelous 17,  
10 433, Athens, Greece

intercalating agent capable of inhibiting both topoisomerases I and II [7–10] and is in phase II clinical trial. For the 9-aminoacridine class of these compounds, the parent of which is 9-amino-DACA, there are tight correlations between ligand structure, cytotoxicity and DNA-binding kinetics [11–15]. Some of these compounds have been determined by X-ray crystallography. In the crystal structures of 9-amino-DACA and its 5-fluoro derivative bound to the hexanucleotide d(CGTAACG)<sub>2</sub> [16, 17], and of 6-Br-9-amino-DACA bound to the brominated hexanucleotide d(CG<sup>5Br</sup>UACG)<sub>2</sub> [18], the 4-carboxamide group lies in the plane of the chromophore, and the carbonyl oxygen atom forms an internal hydrogen bond with the protonated N10 nitrogen of the acridine ring. The side chain lies in the DNA major groove with its protonated *N,N*-dimethylamino group forming hydrogen-bonding interactions with the O6 and N7 atoms of guanine G2. In each case, a hydrogen-bonded water molecule bridges the NH of the carboxamide group to the guanine G2 phosphate at the intercalation site. These structures have provided a molecular rationale for understanding the structure-activity relationships for anti-tumor activity and enabled a mechanistic interpretation of the dependence of kinetics on ligand structure [16, 17]. In particular, they have permitted the critical step in the dissociation kinetics profile that correlates with cytotoxicity and anti-tumor activity to be identified with the side chain-guanine interaction [15–17]. Attempts to crystallize DACA, its phenazine analogue and related agents with d(CGTAACG)<sub>2</sub> have yielded an unusual quadruplex which does not add to our understanding of their complexes with duplex DNA [19–21], and other DNA sequences have failed to give diffracting crystals.

The most significant features of the structure-activity relationships are that a nitrogen atom must be *peri* to the carboxamide, that the carboxamide must have an unsubstituted NH group, and that there must be two methylene groups between the carboxamide NH and the terminal protonated *N,N*-dimethylamino group [11, 16]. To the best of our knowledge there are only limited examples in the recent bibliography regarding the exploration of the energy consequences between intercalative ligands and oligonucleotides. Theoretical investigation of the  $\pi$ - $\pi$  interactions between rhodomycin B and the guanine-cytosine base pair by molecular mechanics and *ab initio* molecular-orbital techniques is referred by Setzer et al. [22], whereas the intermolecular interaction between alkaloids such staurosporine and skimmianine and guanine-cytosine by *ab initio* molecular techniques has also been reported by Byler et al. [23]. Furthermore, semiempirical calculations carried out by Loza-Mejía et al. regarding the interaction between 9-anilinoacridines, 9-anilinothiazolo[5,4-*b*]quinolines and 9-anilinoimidazo[4,5-*b*]quinolines and selected hexanucleotides revealed the strong influence of the replacement of the

benzene moiety for a heterocyclic ring on the simulation of DNA-intercalator complexes [24]. Also, molecular dynamic techniques were used in order to examine energetic interactions between anti-tumor mono- or bisnaphthalimides and d(TG)<sub>2</sub> and d(ATGCAT)<sub>2</sub> [25, 26]. Our aim in this molecular modeling study was to discover whether the *N,N*-dimethylaminoethyl or *N,N*-dimethylaminobutyl side chain of a range of intercalated tricyclic carboxamides with neutral chromophores can form hydrogen-bonding interactions with the O6/N7 atoms of guanine in a manner similar to that of 9-amino-DACA. We believe that our findings could be useful for the design of new chromophores with better DNA-binding activity and anti-tumor action. Furthermore, we think it will be interesting to explore the interaction of these chromophores with DNA, since these kind of molecules are used as dyes in many materials (cosmetics, drugs and food) consumed by people.

Thus, in the inadequacy of direct structural information, and in light of some studies of kinetics of dissociation of DNA-ligand complexes and the known similarities in the DNA binding characteristics of DACA and 9-amino-DACA, we constructed computer models of the complexes of 9-amino-DACA and a range of related linear cyclic carboxamides (compounds 1–12, Fig. 1) [6, 27, 28] with oligonucleotides d(CCGGCGCCGG)<sub>2</sub> and d(CGCGAATTCGCG)<sub>2</sub>. We examined for comparison reasons quinoline-8-carboxamide, a bicyclic system with *N peri* to carboxamide side chain, which is known to bind to DNA in a non-intercalative fashion and is inactive as an anti-tumor agent (Fig. 1) [29].

Since our results suggested that the side chains of systems which bear a *peri* nitrogen atom (4–7) make interactions with DNA in a manner similar to the side chain of 9-amino-DACA (1), we performed also molecular dynamics simulations at 298 K for 15 ns for the complexes of compounds 1 and 4–7 with d(CCGGCGCCGG)<sub>2</sub> and d(CGCGAATTCGCG)<sub>2</sub>.

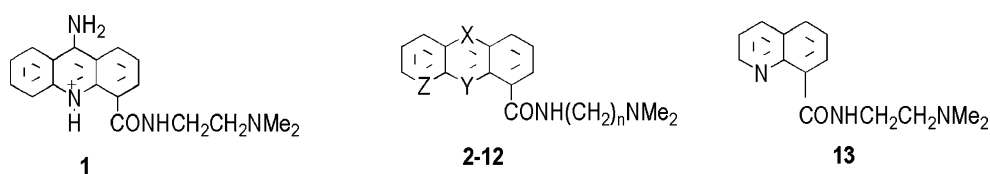
## Computational methods

### Molecular mechanics and molecular dynamic simulations

Insight into the mode of binding of compounds 1–13 to DNA was gained through molecular modeling, molecular mechanics and dynamics techniques using the oligomers d(CCGGCGCCGG)<sub>2</sub> and d(CGCGAATTCGCG)<sub>2</sub>.

The sequence was selected according to the preference for intercalation at 5'-CpG-3' steps exhibited by mono-intercalating acridine derivatives to DNA and to modeling experience [15–17, 30–35] with related intercalating agents.

Ring systems 1–13 were model-built in Hyperchem 7.5 [36] and Spartan 04 program packages [37] using standard

**Fig. 1** Structures of compounds studied

- 2:** X,Y,Z=CH, n=2  
**3:** X=N, Y,Z=CH, n=2  
**4:** X,Z=CH, Y=N, n=2  
**5:** X=CH, Y,Z=N, n=2  
**6:** X,Y=N, Z=CH, n=2  
**7:** X,Y,Z=N, n=2  
**8:** X=NO, Y,Z=N, n=2  
**9:** X=O, Y=NH, Z=CH, n=2  
**10:** X=CO, Y=O, Z=CH, n=2  
**11:** X,Z=CH, Y=N, n=4  
**12:** X,Y=N, Z=CH, n=4

geometries, which were fully optimized by means of the program Spartan 04 with Hartree-Fock method and the 6-31 G\* basis set (Fig. 2).

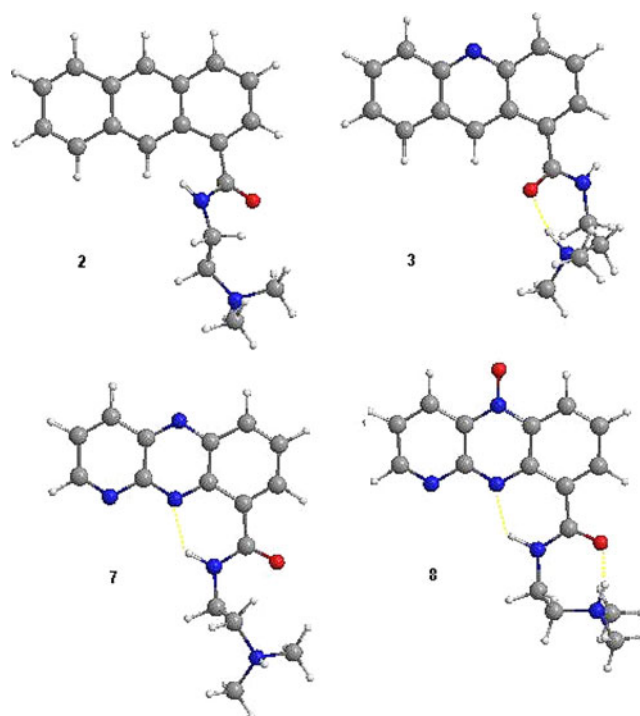
The crystal structures of oligonucleotides d(CCGGCGCCGG)<sub>2</sub> (PDB:1cgc) and d(CGCGAATTCGCG)<sub>2</sub> (PDB: 1d30) were downloaded from the Protein Data Bank (PDB) using Yasara 10.5.14 [38]. The molecules of water and ligand were deleted.

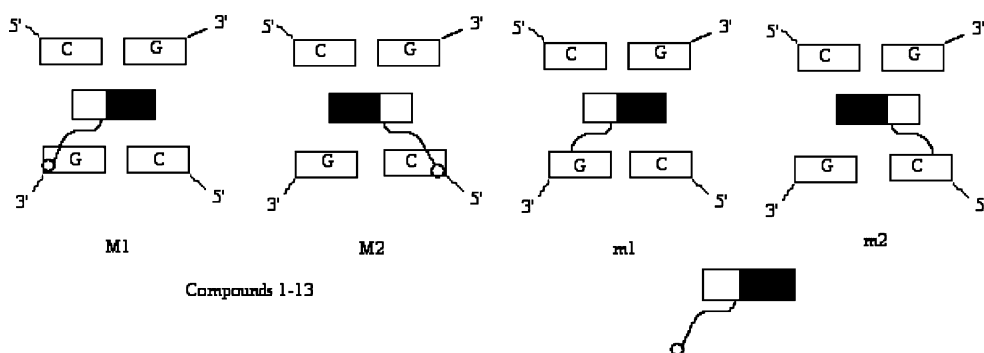
Thirty six model complexes were constructed for compounds 1–3 and 8–13 by inserting the chromophores into the central CG of the d(CCGGCGCCGG)<sub>2</sub> in the four possible orientations relative to the base pairs, denoted M1, M2, m1, and m2, as shown in Fig. 3. Two through the major groove: one with the ring bearing the side chain between the central CG and the other with the ring bearing the side chain between the central GC; and the same orientations leaving the side chain in the minor groove. It is noteworthy that the carboxamide group of compounds with *peri* nitrogen (4–7), can lie in two orientations in the plane of the chromophore, viz, with the carbonyl oxygen pointing toward the *peri* nitrogen, the *cis* position, or away, the *trans* position. In these conformations the resonance stabilization between carboxamide and chromophore is maximized, and in the *trans* position there is the added possibility that the carboxamide NH group can form an internal hydrogen bond with the *peri* nitrogen. Using the mechanical program Spartan 04 and the 6-31 G\* basis set, we calculated that the *trans* conformation of these compounds is more stable than *cis* by about 12.5 kcalmol<sup>-1</sup>.

Therefore, 16 model complexes were constructed for compounds 4–7 with the carboxamide group in *trans* orientation and 16 model complexes were constructed by inserting the chromophores into the d(CCGGCGCCGG)<sub>2</sub> with the ring bearing the side chain stacked between the central CG and the carboxamide groups of the optimized intercalators rotating by 180° (*cis* orientation).

Additional 13 model complexes were constructed for compounds 1–13 by inserting the chromophores into the d(CGCGAATTCGCG)<sub>2</sub> through the major groove, with the ring bearing the side chain stacked between the terminal CG (M1) and four model complexes were constructed for compounds 4–7 with the carboxamide groups of the optimized intercalators rotating by 180° (*cis* orientation).

Processing of data and simulations of all models (1–13) were done using the program Yasara, with the Yamber 3

**Fig. 2** Structures of compounds 2, 3, 7 and 8 after geometry optimization by Hartree-Fock /6-31G\* calculations. Dotted lines indicate hydrogen bonds



**Fig. 3** Schematic view from the major groove of the four complexes between compounds **1–13** and the central CG of  $d(\text{CCGGCGCCGG})_2$ . The white square in the chromophore represents the orientation of the ring with the side chain, and the sphere represents the protonated dimethylamino group from the side chain. Left: complexes with the

linker in the major groove and the ring with the side chain stacked between CG (**M1**), or between GC (**M2**). Right: complexes with the linker in the minor groove and the ring with the side chain stacked between CG (**m1**), or between GC (**m2**)

force field [39]. Van der Waals pairs cutoff distance was 7.86 Å and particle mesh Ewald (PME) long range electrostatics were employed [40]. Simulation structures were energy-minimized to remove bumps and correct the covalent geometry. After removal of conformational stress by steepest descent minimization, the procedure continued by simulated annealing (time step 2 fs, atom velocities scaled down by 0.9 every  $10^{\text{th}}$  step) until convergence was reached, i.e., the energy improved by less than 0.1% during 200 steps.

Periodic boundary simulations were done on orthorhombic cells of extra extension along each axis of the complex of 5 Å. Yasara program assigned force field parameters, filled the cell with water and placed  $\text{Na}^+$  and  $\text{Cl}^-$  counter ions at the most favorable positions to make the cell neutral. The simulation cell was filled with water to a density of 0.997 g/l, and the minimization procedure was repeated, first with the water solvent, then with the oligonucleotide, the ligand, and finally with the whole system. Minimization was followed by a short equilibration procedure to 298 K. Resulting minimized and equilibrated models were subsequently used for MD simulations. Analysis of data was also done with Yasara, using the distributed computing facility. Data is presented as mean  $\pm$  standard deviation.

Trajectories of MD simulations were sampled at intervals of 1 ps, for a total of 15 ns for each model (Fig. 4). We analyzed the stability of trajectories to ensure that the models were at equilibrium. The MD trajectories were inspected, also visually, to assess their structural stability. The gyration radius of the models is maintained quite constant throughout all the trajectories (average values of 12.3–12.9 $\pm$ 0.1–0.3 Å). The quantitative assessment of the stability of the trajectory has been done by measuring two parameters: the root mean square (RMS) of the complexes, and the average potential energy of the system. These two parameters should not vary considerably during the simulation when the system is at equilibrium. During the

first 200 ps there is a change of energy due to the relaxation of the system after assignment of initial velocities. The potential energy for all trajectories converges to a constant average value, demonstrating the reproducibility of the simulations. We considered that the systems reached equilibrium after 500 ps equilibration, as they reached mean energy values that were less than 3% different from the average asymptotic values. Hence, all average values presented were evaluated after the first 700 ps of simulation. The root-mean-square deviation of the complexes of compounds **1** and **4–7** (in *trans* and *cis* orientation) with  $d(\text{CCGGCGCCGG})_2$  and  $d(\text{CGCGAATTCGCG})_2$  with respect to the initial structures were evaluated. The RMSD values ranged from 0.67 to 1.2 (for the *cis* orientation) $\pm$ 0.1–0.4 Å (available as supporting information, Fig. 5). The potential energy of the system was defined as the average force field energy of measured after the first 700 ps simulation in all trajectories (available as supporting information, Fig. 6). The potential energy values ranged from  $-45130$  (**1**) to  $-34716$  [(**5**) in the *cis* conformation] $\pm$ 86–124 kcalmol $^{-1}$ .

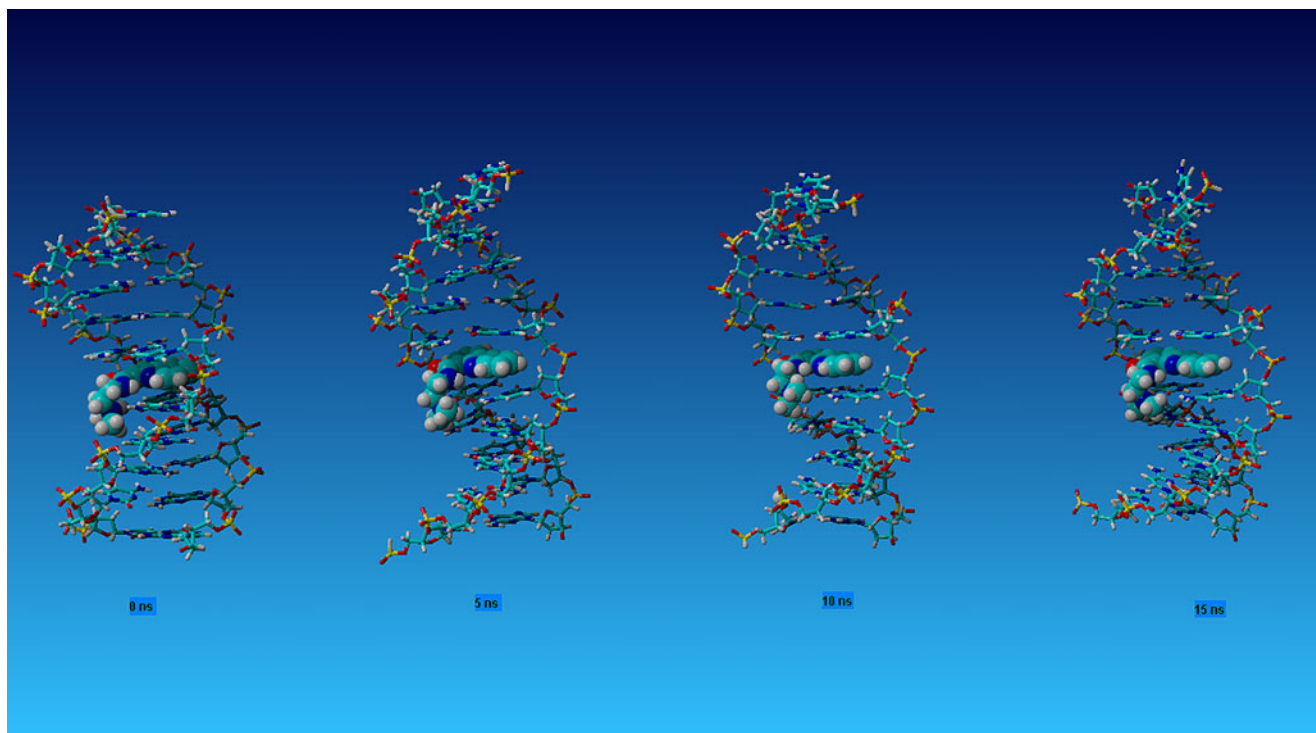
## Results

### Energy minimization

Results are summarized in Table 1.

The model compounds studied can be classified with reference to calculated internal hydrogen bonds, as follows:

- A. Compounds **1**, **4–8**, and **11–13**, which have a  $-\text{NH}^+$  group or  $-\text{N}$  atom *peri* to the carboxamide. These may be divided into two types.
  1. Compounds **1**, and **4–7**, which have one internal hydrogen bond between the carboxamide and the  $-\text{NH}^+$  or  $-\text{N}$  group in the *peri* position.



**Fig. 4** Graphic representation of DACA-CCGGCGCCGG model in the initial state, and after 5,10 and 15 ns simulation. The figures show that the structure has remained stable for the duration of the simulation, without incurring large conformational changes

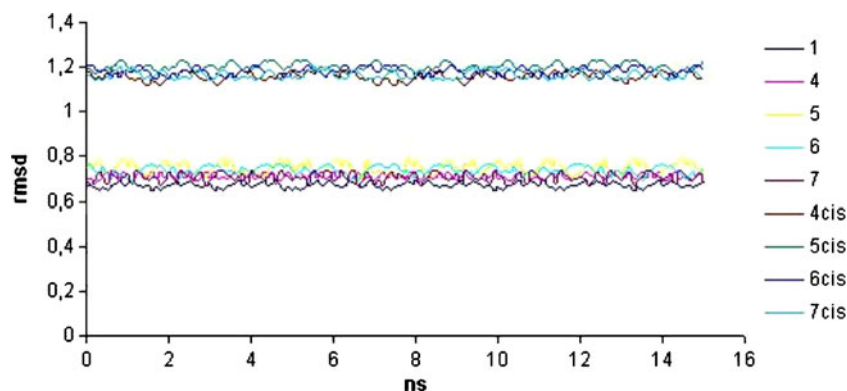
2. Compounds **8**, **11–13**, which have an additional internal hydrogen bond between the carboxamide and the charged amino group.
- B. Compounds **2**, **3**, **9** and **10**, which lack a nitrogen *peri* to the carboxamide. These may be divided into three types.
1. Compound **2**, which lacks internal hydrogen bonds.
  2. Compound **3**, which has one internal hydrogen bond between the carboxamide and the dimethylammonium group.
  3. Compounds **9**, and **10**, which have two internal hydrogen bonds : one between the carboxamide and the –NH or –O group in the *peri* position and the other between the carboxamide and the dimethylammonium group.

In the complexes of compounds **1** and **4–7** with the oligonucleotide d(CCGGCGCCGG)<sub>2</sub> in the four possible orientations, the *N,N*-dimethylammonium group forms strong hydrogen bonding interaction with the G6-N7/O6 atoms, O6 of G16, O2 of C5 and O2 of C15 for the M1, M2, m1 and m2 orientations, respectively.

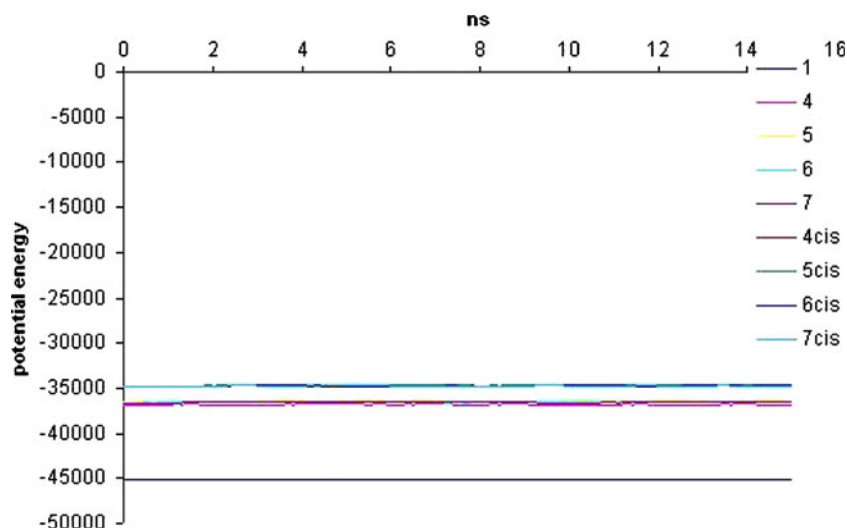
In the four (2)-CCGGCGCCGG complexes, the *N,N*-dimethylammonium group does not form a hydrogen bond with the heteroatoms of the base pairs in the M1 orientation, whereas the carboxamide group hydrogen bonds strongly to O6 of G16, N2 of G16 and N3 of G6 for the M2, m1 and m2 orientations, respectively.

In the (3)-CCGGCGCCGG complexes the *N,N*-dimethylammonium group does not form a hydrogen bond with the base pairs in the M1 orientation, whereas the carbox-

**Fig. 5** Plots of the variation of the rmsd of compounds **1** and **4–7** throughout the simulation



**Fig. 6** Plots of the variation of the potential energy of compounds **1** and **4–7** throughout the simulation



amide group bonds strongly to N3 of C15, the sugar O4 atom of G6 and the sugar O4 atom of G16 for the M2, m1 and m2 orientations, respectively.

In the complexes of compounds **8** and **11–13** with the decanucleotide d(CCGGCGCCGG)<sub>2</sub> in the four possible orientations, the carboxamide and the *N,N*-dimethylammonium groups are not sufficiently close to any electronegative base pair atom to be involved in hydrogen bonding. Additionally, in the M1 orientation the H atom of the carboxamide bonds with the *peri* heteroatom of the chromophore and the O atom of the carboxamide bonds with the *N,N*-dimethylammonium group (compounds **11** and **13**).

In the four (**9**)-CCGGCGCCGG complexes, the *N,N*-dimethylammonium group makes strong hydrogen bonding interaction with the N4 of C5, N3 of G6 and G16 for the M1, m1 and m2 orientations, respectively. On the contrary, there is no hydrogen bonding interaction between *N,N*-dimethylammonium group and the base pairs in the M2 orientation. The O atom of the carboxamide bonds with the *N,N*-dimethylammonium group and with the H atom of the NH of the chromophore in the M1 orientation.

In the four (**10**)-CCGGCGCCGG complexes, the *N,N*-dimethylammonium group hydrogen bonds to N3 of G6 and makes a weak interaction with the N3 of G16 for the m1 and m2 orientations, respectively. In the M1 orientation, there is no hydrogen bonding interaction between *N,N*-dimethylammonium group and the base pairs, whereas there is hydrogen bonding interactions between the H atom of the carboxamide group and the *peri* heteroatom of the chromophore and additionally between the O atom of the carboxamide moiety and the *N,N*-dimethylammonium group.

#### Molecular dynamic simulations

In the molecular dynamic simulation of the (**1**)-CCGGCGCCGG complex, the hydrogen of the *N,N*-dime-

thylammonium is strongly hydrogen bonded to O6 of guanine-6 and makes a weak interaction with the N7 of guanine-6. In the molecular dynamic simulation of the (**1**)-CGCGAATTCGCG complex, the hydrogen of the *N,N*-dimethylammonium makes a strong hydrogen bond solely to O6 of guanine-2. The side-chain torsion angle shows a *gauche* conformation relating the two side-chain nitrogen atoms, just as it occurs in the crystal structures [17].

In the molecular dynamic simulation of the (**4**)-CCGGCGCCGG complex, with the carboxamide group in the *cis* orientation (the carbonyl oxygen points toward the *peri* nitrogen), the hydrogen of the *N,N*-dimethylammonium moiety makes strong hydrogen bonding interaction with the O6 of guanine-6 and feebly interacts with the N7 of the same guanine. In the (**4**)-CCGGCGCCGG complex, with the carboxamide group in the *trans* orientation (the carbonyl oxygen points away the *peri* nitrogen), the hydrogen of the *N,N*-dimethylammonium moiety makes strong hydrogen bonding interaction with the O6 of guanine-6.

In the molecular dynamic simulation of the (**4**)-CGCGAATTCGCG complex, with the carboxamide group in the *cis* orientation, the hydrogen of the *N,N*-dimethylammonium moiety makes a strong hydrogen bond solely with the O6 of guanine-2. In the (**4**)-CGCGAATTCGCG complex, with the carboxamide group in the *trans* orientation, the hydrogen of the *N,N*-dimethylammonium hydrogen strongly bonds to the O6 of guanine-2 and weakly interacts with the N7 of guanine-2.

In the molecular dynamic simulation of the (**5**)-CCGGCGCCGG complex, with the carboxamide group in the *cis* orientation, the hydrogen of the *N,N*-dimethylammonium moiety forms a hydrogen bond with the O6 of guanine-6 and is close to G6-N7, with distance H(N)...O (G) of 3.2 Å ± 0.3. In the (**5**)-CCGGCGCCGG complex, with the carboxamide group in the *trans* orientation, the

**Table 1** Hydrogen bonds\* of a) N, N-dimethylammonium group of compounds 1–13 with Guanine-O6/N7 or the carboxamide moiety and b) the carboxamide moiety with chromophore

No	Compound	peri	n	Orientation	Oligonucleotides	Hydrogen bond of the N, N-dimethylammonium group with Guanine-O6/N7	Hydrogen bond of the carboxamide O with NH <sup>+</sup> or NH <sup>‡</sup>	Hydrogen bond of the carboxamide H with <i>peri</i> heteroatom <sup>‡</sup>	Hydrogen bond of the carboxamide O with N,N-dimethylammonium group <sup>‡</sup>	Interaction with Guanine <sup>†</sup>
1	Amino DACA	NH <sup>+</sup>	2	cis	d(CCGGGCCGG) <sub>2</sub> / d(CCGGAATTCGG) <sub>2</sub>	Yes (G6/G2)	Yes	No	No	Yes
2	Anthracene	CH	2	-	d(CCGGGCCGG) <sub>2</sub>	No	-	No	No	No
3	Acridine-1-carboxamide	CH	2	-	d(CCGGGCCGG) <sub>2</sub>	No	-	No	Yes	No
4	Acridine-4-carboxamide (DACA)	N	2	cis/trans	d(CCGGGCCGG) <sub>2</sub> / d(CCGGAATTCGG) <sub>2</sub>	Yes (G6/G2)	-	Yes	No	Yes
5	Azaeridine	N	2	cis/trans	d(CCGGGCCGG) <sub>2</sub> / d(CCGGAATTCGG) <sub>2</sub>	Yes (G6/G2)	-	Yes	No	NT
6	Phenazine	N	2	cis/trans	d(CCGGGCCGG) <sub>2</sub> / d(CCGGAATTCGG) <sub>2</sub>	Yes (G6/G2)	-	Yes	No	Yes
7	Pyridoquinoxaline	N	2	cis/trans	d(CCGGGCCGG) <sub>2</sub> / d(CCGGAATTCGG) <sub>2</sub>	Yes (G6/G2)	-	Yes	No	NT
8	Oxopyridoquinoxaline	N	2	cis/trans	d(CCGGGCCGG) <sub>2</sub>	No	-	Yes	Yes	NT
9	Phenoxazine	NH	2	cis	d(CCGGGCCGG) <sub>2</sub>	No	Yes	No	Yes	No
10	Xanthenone	O	2	cis/trans	d(CCGGGCCGG) <sub>2</sub>	No	-	Yes	Yes	No
11	Acridine-4-carboxamide	N	4	cis/trans	d(CCGGGCCGG) <sub>2</sub>	No	-	Yes	Yes	No
12	Phenazine	N	4	cis/trans	d(CCGGGCCGG) <sub>2</sub>	No	-	Yes	Yes	No
13	Quinoline	N	2	cis/trans	d(CCGGGCCGG) <sub>2</sub>	No	-	Yes	Yes	NT

\*External hydrogen bonds obtained by inserting the chromophore in the M1 orientation relative to the base pairs ‡ Intermolecular hydrogen bonds, † kinetic study by Wakelin et al., NT = Not tested

protonated *N,N*-dimethylamino group makes a strong hydrogen bond to O6 of guanine-6 and weakly interacts with the N7 of guanine-6. In the (5)-CGCGAATTCGCG complex, with the carboxamide group in the *cis* orientation, the protonated *N,N*-dimethylamino group forms a hydrogen bond with O6 of guanine-2. In the (5)-CGCGAATTCGCG complex, with the carboxamide group in the *trans* orientation, the *N,N*-dimethylammonium moiety forms strong hydrogen bonding interaction with O6 of G2 and is also in contact of  $3.3 \text{ \AA} \pm 0.2$  with N7 of G2.

In the molecular dynamic simulation of the (6)-CCGGCGCCGG complex, with the carboxamide group in the *cis* orientation, the protonated *N,N*-dimethylamino group strongly hydrogen bonds to G6-O6 and is close to G6-N7, with distance H(N)...N(G) of  $3.4 \text{ \AA} \pm 0.2$ . In the same complex, with the carboxamide group in the *trans* orientation, the *N,N*-dimethylammonium group forms two hydrogen bonds to G6-N7/O6 atoms, a weak and a strong, with distances of  $3.3 \pm 0.2$  and  $2.1 \text{ \AA} \pm 0.2$ , respectively.

In the molecular dynamic simulation of the (6)-CGCGAATTCGCG complex, with the carboxamide group in the *cis* orientation, the protonated *N,N*-dimethylamino group forms a strong hydrogen bond to G2-O6. In the same complex, with the carboxamide group in the *trans* orientation, the protonated *N,N*-dimethylamino group forms a strong hydrogen bond to G2-O6 and is in close contact with G2-N7, with distance H(N)...N(G) of  $3.6 \text{ \AA} \pm 0.3$ .

In the molecular dynamic simulation of the (7)-CCGGCGCCGG complex, with the carboxamide group in the *cis* orientation, the protonated *N,N*-dimethylamino group makes strong hydrogen bonding interaction with G6-O6 and is close to G6-N7, with distance H(N)...O(G) of  $3.8 \text{ \AA} \pm 0.2$ . In the same complex, with the carboxamide group in the *trans* orientation, the protonated *N,N*-dimethylamino group strongly hydrogen bonds to G6-O6, with distance H(N)...O(G) of  $1.9 \text{ \AA} \pm 0.2$ .

In the molecular dynamic simulation of the (7)-CGCGAATTCGCG complex, with the carboxamide group in the *cis* orientation, the *N,N*-dimethylammonium group gives a good hydrogen bond to G2-O6, with distance H(N)...N(G) of  $2.1 \text{ \AA} \pm 0.3$ . In the same complex, with the carboxamide group in the *trans* orientation, the protonated *N,N*-dimethylamino group makes a strong hydrogen bond solely to O6 of guanine-2, with distance H(N)...O(G) of  $1.9 \text{ \AA} \pm 0.1$ .

## Discussion

We have found by molecular mechanics methods that in the, phenazine, azacridine, pyridoquinoxaline and acridine-4-carboxamide series **1**, **4–7** the *N,N*-dimethylaminoethyl side chain does interact with guanine of two oligonucleo-

tides in M1 orientations, with the 4-carboxamide group rotated *cis* or *trans* with respect to the ring nitrogen. By contrast, when the side chain is *peri* to a CH group, as in the anthracene and acridine-1-carboxamide compounds, **2** and **3**, the side chain fails to make this interaction. An important difference between two ligand classes is that, like 9-amino-DACA, compound **1**, the carboxamide groups of compounds **4–7** can lie in the plane of the chromophore, whereas in the case of the anthracene and acridine-1-ligands, steric interaction with the *peri* hydrogen atom prevents the carboxamide from being coplanar. Thus, it is obvious that coplanarity of the carboxamide and chromophore is a prerequisite for effective side chain-guanine interactions among the linear tricyclic carboxamides such as DACA and its analogues.

However, when the distance between the 4-carboxamide and *N,N*-dimethylamino group is lengthened (compounds **11** and **12**), the interaction with guanine is lost probably due to the internal hydrogen bond between the carbonyl oxygen and the protonated *N,N*-dimethylamino group.

In the cases of the oxopyridoquinoxaline and quinoline ligands, **8** and **13**, the existence of the internal hydrogen bond between the carbonyl oxygen and the protonated *N,N*-dimethylamino group seems to prevent the *N,N*-dimethylaminoethyl side chain from interacting with guanine.

In the case of the phenoxazine ligand **9**, the carbonyl oxygen forms an internal hydrogen bond in the *cis* configuration with the *peri* NH atom, but the existence of a second internal hydrogen bond between the carbonyl oxygen and the protonated *N,N*-dimethylamino group seems to prevent the *N,N*-dimethylaminoethyl side chain from interacting with guanine. Clearly, for this ligand the *trans* carboxamide orientation is prohibited by steric collision of the two NH groups, which will force an angular conformation between chromophore and carboxamide. In the case of the xanthenone ligand **10**, the NH of the carboxamide forms an internal hydrogen bond in the *trans* configuration with the *peri* oxygen atom, but the existence of a second internal hydrogen bond between the carbonyl oxygen and the protonated *N,N*-dimethylamino group also seems to prevent the *N,N*-dimethylaminoethyl side chain from interacting with guanine. Owing to this internal hydrogen bond, the *cis* xanthenone carboxamide configuration should be unable to bond successfully with the guanine O6/N7 atoms.

From an assessment of the above data regarding tricyclic carboxamides, prerequisite for the interaction between protonated *N,N*-dimethylaminoethyl moiety and guanine is the formation of only one internal hydrogen bond between carboxamide and *peri* NH+ in the case of 9-amino-DACA (**1**) or *peri* N in the cases of DACA (**4**), azacridine (**5**), phenazine (**6**) and pyridoquinoxaline (**7**). The presence of an additional internal hydrogen bond between oxygen



carboxamide and protonated *N,N*-dimethylamino group in the cases of tricyclic systems bearing *peri* NH (phenoxazine, **9**) or O (xanthenone, **10**) group, prevents the interaction between side chain and guanine. Also, the formation of one internal hydrogen bond between oxygen carboxamide and protonated *N,N*-dimethylamino group inhibits the interaction between side chain and guanine in the case of acridine-1-carboxamide (**3**). It is remarkable, that our findings correlate well with reported results obtained from the kinetic studies of the binding of such tricyclic carboxamides to DNA. According to Wakelin et al. the complexes of **1**, **4** and **6** dissociate from calf thymus DNA by a kinetic pathway involving four discernible steps, indicating that they interact favorably with the guanine O6/N7 atoms. By contrast, the tricyclic carboxamides **2**, **3**, **9**, **10**, **11** and **12** with neutral chromophore dissociate from DNA by a different mechanism in which it appears their side chain fail to interact with guanine. There is no data available in the literature regarding the kinetics of compounds **5**, **7**, **8** and **13** with DNA and therefore correlation with our results is not possible.

In all the examined complexes of compounds **1** and **4–7** with the oligonucleotides, the distance between the charged amino group and O6 guanine base is rather constant, i.e., from 1.8 to 2.2 Å, indicating the presence of a hydrogen bond that remains stable during the entire molecular dynamic simulation. The deviations in kinetic, potential energy and in the angles of hydrogen bonds were small. Based on the above observations, we can assume that the complexes do not experience large conformational changes during the sampling time with respect to the initial structure.

Finally, based on the literature [6] and our molecular modeling experiments we can assume that there is a broad positive correlation between cytotoxicity of the protonated *N,N*-dimethylamino carboxamides with neutral chromophores and their interaction to guanine, with the compounds **4**, **5** and **6** with only one internal hydrogen bond between carboxamide and *peri* nitrogen being potent cytotoxins. The compounds **11** and **12** (with two internal hydrogen bonds: one between carboxamide and *peri* nitrogen and the second between carbonyl oxygen and protonated *N,N*-dimethylamino group) are also potent cytotoxins. The compounds **2** (with no internal hydrogen bonds), **3** (with one internal hydrogen bond between carbonyl oxygen and protonated *N,N*-dimethylamino group) and the compounds **9** and **10** (with two internal hydrogen bonds: one between carboxamide and *peri* heteroatom and the second between carbonyl oxygen and protonated *N,N*-dimethylamino group) are less cytotoxic. The compound **13** (with two internal hydrogen bonds: one between carboxamide and *peri* heteroatom and the second between oxygen carboxamide and protonated *N,N*-dimethylamino group) is inactive. In

conclusion, prerequisite for the interaction between protonated *N,N*-dimethylaminoethyl moiety and guanine is the formation of only one internal hydrogen bond between carboxamide and *peri* NH<sup>+</sup> in the case of 9-amino-DACA or *peri* N in the cases of DACA, azacridine, phenazine and pyridoquinoxaline. The presence of an additional internal hydrogen bond between oxygen carboxamide and protonated *N,N*-dimethylamino group in the cases of tricyclic systems bearing *peri* NH or O group, prevents the interaction between side chain and guanine and leads to reduction of the cytotoxicity. Additionally, the present molecular dynamics simulations show that the compounds **1** and **4–6** are capable of forming rather stable DNA complexes with small deviations in potential energy, RMSD and in the angles of hydrogen bonds, which is in accordance with good *in vitro* activities, further supported by *in vivo* experiments.

## References

1. Varvaresou A, Tsoinias A, Papadaki-Valiraki A, Siatra-Papastaiakoudi TH (1996) *Bioorg Med Chem Lett* 6:861–864
2. Kapadia GJ, Tokuda H, Shridhar R, Balasubramanian V, Takayasu J, Bu P, Enjo F, Takasaki M, Konoshima T, Nishino H (1998) *Cancer Lett* 129:87–95
3. Varvaresou A, Iakovou K, Gikas E, Fichtner I, Fiebig HH, Kelland LR, Double JA, Bibby MC, Hendriks HR (2004) *Anticancer Res* 24:907–920
4. Varvaresou A, Tsirivas E, Tsaoula E, Protopapa E (2005) *Rev Clin Pharmacol Pharmacokin* 19:11–16
5. Miller KJ, Newlin DD (1982) *Biopolymers* 21:633–652
6. Wakelin LPG, Adams A, Denny WA (2002) *J Med Chem* 45:894–901
7. Atwell GJ, Rewcastle GW, Baguley BC, Denny WA (1987) *J Med Chem* 30:664–669
8. Schneider E, Darkin SJ, Lawson PA, Ching LM, Ralph RK, Baguley BC (1988) *Eur J Cancer Clin Oncol* 24:1783–1790
9. Baguley BC, Zhuang L, Marshall E (1995) *Cancer Chemother Pharmacol* 36:244–248
10. Finlay GJ, Riou JF, Baguley BC (1996) *Eur J Cancer A* 32A:708–714
11. Atwell GJ, Cain BF, Baguley BC, Finlay GJ, Denny WA (1984) *J Med Chem* 27:1481–1485
12. Denny WA, Roos IAG, Wakelin LPG (1986) *Anticancer Drug Des* 1:141–147
13. Rewcastle GW, Atwell GJ, Chambers D, Baguley BC, Denny WA (1986) *J Med Chem* 29:472–477
14. Denny WA, Atwell GJ, Rewcastle GW, Baguley BC (1987) *J Med Chem* 30:658–663
15. Wakelin LPG, Atwell GJ, Rewcastle GW, Denny WA (1987) *J Med Chem* 30:855–861
16. Adams A, Guss JM, Collyer CA, Denny WA, Wakelin LPG (1999) *Biochemistry* 38:9221–9233
17. Adams A, Guss JM, Collyer CA, Denny WA, Prakash AS, Wakelin LPG (2000) *Mol Pharmacol* 58:649–658
18. Todd AK, Adams A, Thorpe JH, Denny WA, Wakelin LPG, Cardin CJ (1999) *J Med Chem* 42:536–540
19. Thorpe JH, Hobbs JR, Todd AK, Denny WA, Charlton P, Cardin CJ (2000) *Biochemistry* 39:15055–15061
20. Adams A, Guss JM, Collyer CA, Denny WA, Wakelin LPG (2000) *Nucleic Acids Res* 28:4244–4253

21. Yang XI, Robinson H, Gao YG, Wang AHJ (2000) *Biochemistry* 39:10950–10957
22. Setzer WN, Rozmus GF, Setzer MC, Schmidt JF, Vogler B, Reeb S, Jackes BR, Irvine AK (2006) *J Mol Model* 12:703–711
23. Byler KG, Wang C, Setzer WN (2009) *J Mol Model* 15:1417–1426
24. Loza-Mejía MA, Castillo R, Lira-Rocha A (2009) *J Mol Graph Model* 27:900–907
25. Braña MF, Cacho M, García MA, Pascual-Tereza B, Ramos A, Acero N, Llinares F, Muñoz-Mingarro D, Abradelo C, Rey-Stolle MF, Yuste M (2002) *J Med Chem* 45:5813–5816
26. Braña MF, Cacho M, García MA, Pascual-Tereza B, Ramos A, Dominguez MT, Pozuelo JM, Abradelo C, Rey-Stolle MF, Yuste M, Báñez-Coronel M, Lacal JC (2004) *J Med Chem* 47:1391–1399
27. Chen Q, Deady LW, Baguley BC, Denny WA (1994) *J Med Chem* 37:593–597
28. Varvaresou A, Iakovou K (2002) *J Heterocycl Chem* 39:1173–1176
29. Atwell GJ, Bos CD, Baguley BC, Denny WA (1988) *J Med Chem* 31:1048–1052
30. Müller W, Crothers DM, Thara T (1975) *Eur J Biochem* 54:267–277
31. Bailly C, Denny WA, Mellor LE, Wakelin LPG, Waring MJ (1992) *Biochemistry* 31:3514–3524
32. Fischer G, Pindur U (1999) *Pharmazie* 54:3–93
33. Wadkins RM, Graves DE (1991) *Biochemistry* 30:4277–4283
34. Krugh TR (1994) *Curr Opin Struct Biol* 4:351–364
35. Krugh TR, Reinhardt CG (1975) *J Mol Biol* 97:133–162
36. Hyperchem, Release 7.5 (2003) Hypercube Inc
37. Spartan 04, 1991–2003, Wavefunction Inc
38. Krieger E, Koraimann G, Vriend G (2002) *Proteins* 47:393–402
39. Krieger E, Darden T, Nabuurs SB, Finkelstein A, Vriend G (2004) *Proteins* 57:678–683
40. Kawata M, Nagashima U (2001) *Chem Phys Lett* 340:165–172